

## Begeleidingsformulier aanvraag dierproef DEC- UM

Versie 2006

Herziene versie

DECNR: 2011-120

Ontvangen: 03-10-2011

|                        |                             |
|------------------------|-----------------------------|
| DEC datum goedkeuring# | Type aanvraag 2             |
|                        | Nieuw / Herz.versie / Pilot |

VROM/GGO  
NR<sup>3</sup>LNV/CBDNR<sup>4</sup>

|              |       |        |                    |      |               |          |         |
|--------------|-------|--------|--------------------|------|---------------|----------|---------|
| Hoofdproject | CARIM | NUTRIM | Hersenen en gedrag | GROW | biomaterialen | Ander UM | Geen UM |
|--------------|-------|--------|--------------------|------|---------------|----------|---------|

|             |          |             |          |          |  |  |  |
|-------------|----------|-------------|----------|----------|--|--|--|
| Deelproject | 1. 2. 3. | 1. 2. 3. 4. | 1. 2. 3. | 1. 2. 3. |  |  |  |
|-------------|----------|-------------|----------|----------|--|--|--|

|                      |  |
|----------------------|--|
| Financieel beheerder |  |
|----------------------|--|

Budgetnummer 3098.2246N

Titel van het onderzoek:

Unraveling the role of micro in a model of heart failure

startdatum 1 oktober 2011

einddatum 9 1 oktober 2015

Duur van de proef<sup>10</sup>: 49 days

|                                                      | Naam                    | Tel (+ Tel privé enkel VO, VVO en VM) | E-mailadres | Bevoegdheid <sup>5</sup> | Cap. groep /afdeling |
|------------------------------------------------------|-------------------------|---------------------------------------|-------------|--------------------------|----------------------|
| 1. Verantwoordelijk onderzoeker (VO)                 |                         |                                       |             | Art.9                    |                      |
| 2. Vervanger VO (VVO)                                |                         |                                       |             | Art.9                    |                      |
| 3. Verantwoordelijk medewerker (VM) GGO <sup>7</sup> |                         |                                       |             |                          |                      |
| 4. overige uitvoerenden                              | Artikel 12 functionaris |                                       |             | Art.12                   |                      |
| 5.                                                   | (PI)                    |                                       |             | Art.9                    |                      |
| 6.                                                   |                         |                                       |             | Art.9                    |                      |
| 7.                                                   |                         |                                       |             |                          |                      |

| Diergroep                 | 1-2      | 3-4      | 5-6      | 7-8      |
|---------------------------|----------|----------|----------|----------|
| ctrl/exp/sham             | sham     | exp      | sham     | exp      |
| Diersoort                 | muis     | muis     | muis     | muis     |
| Stam                      | C57Bl/6  | C57Bl/6  | C57Bl/6  | C57Bl/6  |
| Construct / mutatie ?     | wildtype | wildtype | wildtype | wildtype |
| Herkomst (leverancier) *  | 1        | 1        | 1        | 1        |
| Aantal                    | 36       | 38       | 36       | 38       |
| Geslacht                  | m        | m        | m        | m        |
| Dieren immuuncompetent ?  | ja       | ja       | ja       | ja       |
| Leeftijd/gewicht          | 6-12 wkn | 6-12 wkn | 4-9 wkn  | 4-9 wkn  |
| Doel van de proef *       | 31       | 31       | 31       | 31       |
| Belang van de proef *     | 1        | 1        | 1        | 1        |
| Toxicologisch onderzoek * | 1        | 1        | 1        | 1        |
| Bijzondere technieken *   | 1        | 1        | 1        | 1        |
| Anesthesie *              | 4        | 4        | 4        | 4        |
| Pijnbestrijding *         | 4        | 4        | 4        | 4        |
| Mate ongerief *           | 4        | 4        | 4        | 4        |
| Toestand dier einde exp*  | 1        | 1        | 1        | 1        |

\* VHL-coderingen zie bijlage

*Application for the conduction of animal experiments (DEC-UM)*

**Title:** Unraveling the role of microRNA-133 in a model of heart failure

**Accountability****1. Aim of the experiment.**

Arterial hypertension is an important risk factor for heart failure (HF), a significant cause of mortality worldwide. But still our knowledge of the mechanisms involved in development of heart failure is incomplete.

We identified miR-133 as differentially regulated in human and mouse heart failure. We suggest that miR-133 has a positive impact on the development of heart failure by repressing the expression of deleterious gene products, one of which is the pro-hypertrophic *ANP*, a known target of miR-133.

We hypothesize that inhibition of miR-133 will aggravate the development of hypertrophy and heart failure, while overexpressing pre-miR-133 will protect against heart failure. We will study these hypotheses in a mouse model of heart failure in which cardiac hypertrophy is induced by angiotensin II infusion. We will administer inhibitors of miR-133 (antagomiRs) or overexpressors of pre-miR-133 (encoded by adeno-associated viral vector type 9, AAV9) via tail vein injection and examine cardiac function and histology.

**2. Social relevance and/or scientific importance**

Heart failure is a progressive and severely invalidating disease with a bad prognosis and no treatment available other than symptomatic therapy. Heart failure is the number 2 cause-of-death in Western society, directly after cancer. Most patients suffering from heart failure die within 5 years after diagnosis. The current study will improve our understanding of the mechanisms that lead to heart failure, and of clinically important actors in this process. This is highly relevant on the social as well as economical level.

**3. Alternatives**

The heart is a complex organ composed of different cell types. One cell type interacts with its surrounding environment. Above that, other cell types (immune cells) infiltrate during heart failure and interact with cells that reside in the heart. This creates a highly complex *in vivo* situation, and no *in vitro* model is able to simulate these complex processes. Hence, it is essential that we examine the heart in the *in vivo* setting.

**4. Ethical consideration**

Heart failure is the number 2 cause of death in The Netherlands and other Western countries; yearly more than 6000 people die of this disease in The Netherlands. Current knowledge about the crucial contributing factors is flawed. This animal experiment will contribute to a better understanding of the causes of heart failure, and eventually to better treatment and diagnosis of patients. Given the complexity of the heart and the benefit offered by this study, we consider it essential to conduct an animal study without underestimating the serious inconvenience of using laboratory animals.

## Science

### 5. Scientific background

The pathogenesis of heart failure is complex, and is characterized by enlargement of cardiomyocytes (hypertrophy), cardiac tissue fibrosis and inflammation. It is known that microRNAs, newly discovered RNA molecules that regulate gene expression, influence hypertrophy and fibrosis. An involvement of miR-1 in this process is suggested by downregulation of this microRNA during human and mouse cardiac hypertrophy and failure. We established the mouse model of heart failure, which is predominantly expressed by cardiomyocytes in the heart (see arrows in inset; myocardium stained blue; nuclei in pink).



In this study we will investigate whether manipulation of miR-1 affects the development of hypertrophy and heart failure in a mouse model of hypertension.

First, we will administer antagomiRs targeting miR-1 (or scrambled control) by tail vein injection on three consecutive days, and then subject mice to angiotensin II-induced cardiac hypertrophy by implantation of osmotic minipumps that deliver AngII subcutaneously. Sham mice will receive an osmotic minipump for continuous subcutaneous saline injection.

Second, we will treat mice with AAV9 overexpressing pre-miR-1 (or scrambled control), which is administered by tail vein injection. AAV9 specifically infects cardiomyocytes. After 3 weeks, to allow pre-miR-1 expression in the heart, mice are treated with AngII as above.

### 6. Scientific review

The current project is independently evaluated and approved by the PI of the department.

## Experimental Animals

### 7. Choice experimental animal

#### 7a. Species, strain / origin / final destination

For the study we will use C57Bl/6J mice from Harlan. The mice will be sacrificed at the end of the experiment to assess heart function, histology and RNA/protein levels in the hearts.

#### 7b. Sex

The sex differences between males and females with heart failure are well-known but insufficiently understood. Therefore, to exclude possible hormonal differences, we will use male mice for the study.

#### 7.c. Number

First, we will study the manipulation of miR by administering an inhibitor (antagomiR) to mice, followed by induction of heart failure by angiotensin II infusion. The antagomiR will be compared to controls that receive a scrambled oligonucleotide. Second, we will study the manipulation of miR by administering AAV9 overexpressing pre-mi or pre-scrambled.

Study end point is echocardiographically assessed cardiac function.

Power calculation:

Formula of L. Sachs:

$$n = 2(z_{\alpha/2} - z_{\pi})^2 \cdot (\sigma/\delta)^2 = F \cdot (\sigma/\delta)^2$$

$$F_{0,80} = 15,7 \text{ for } \alpha = 0,05 \text{ and } \pi = 80\%$$

Based on our experience, the variation coefficient for angiotensin II infusion is 20% and the expected relative impact is 20%. The  $P$  value is  $<0.05$  and power 80%.

$$N = 15.7 \cdot (0.2/0.2)^2 = 15.7$$

Angiotensin II-treatment leads to a peri-operative and post-operative mortality of about 15%.

If 15.7 mice represent 85% of the total group, we would need  $(15.7/85) \cdot 100 = 18.5$  or **19 animals** per group.

We would like the same number of surviving mice in the sham groups. Saline-treatment leads to a peri-operative mortality of 10%, but is not associated with post-operative mortality. Therefore, each sham group will consist of  $(15.7/90) \cdot 100 = 17.4$  mice, or **18 mice**.

| Group                                | Number of animals |
|--------------------------------------|-------------------|
| 1. scrambled antagomiR + s.c. saline | 18                |
| 2. antagomiR + s.c. saline           | 18                |
| 3. scrambled antagomiR + s.c. Ang II | 19                |
| 4. antagomiR + s.c. Ang II           | 19                |
| 5. AAV9-pre-scrambled + s.c. saline  | 18                |
| 6. AAV9-pre-mi + s.c. saline         | 18                |
| 7. AAV9-pre-scrambled + s.c. Ang II  | 19                |
| 8. AAV9-pre-mi + s.c. Ang II         | 19                |
| <b>Total</b>                         | <b>148</b>        |

## Animal Experiments

### 8. Experiment

The following experiments will be conducted:

**Tail vein injection of antagomiRs (groups 1-4).** On day -3 to -1, mice will be injected once a day with a volume of 0.2 ml antagomiRs (scrambled or antagomir dissolved in saline, or saline alone), by tail vein injection. For the injections, the mice will briefly be placed in a heating cage and then in a restrainer.

**Tail vein injection of AAV9 (groups 5-8).** On day -21, mice will be injected once with a volume of 0.2 ml AAV9 (scrambled or in saline), by tail vein injection. For the injections, the mice will briefly be placed in a heating cage and then in a restrainer.

**Angiotensin II or saline infusion with subcutaneous minipump (all groups).** This procedure will be performed according to SOP or 28 days. We know that a dose of 2.5 mg/(kg·day) angiotensin II causes elevation of blood pressure, cardiac hypertrophy and failure in C57Bl/6J mice. Implantation of (saline-filled) minipumps in control animals is necessary, because control animals need to undergo similar stress and surgery to be compared to the AngII treated animals.

**Echocardiography** will be performed following SOP.

**Terminal hemodynamic characterization of the mouse (dP/dt)** will be performed according to SOP.

#### Time schedule groups 1-4:

| Day | -3                                                     | -2 | -1 | 0                                                       | 27-28                     |
|-----|--------------------------------------------------------|----|----|---------------------------------------------------------|---------------------------|
|     | Daily i.v. injection of antagomiR or scrambled control |    |    | Echo and implantation of minipump with Ang II or saline | Echo, dP/dt and sacrifice |

#### Time schedule groups 5-8:

| Day | -21                    | 0                                                       | 27-28                     |
|-----|------------------------|---------------------------------------------------------|---------------------------|
|     | i.v. injection of AAV9 | Echo and implantation of minipump with Ang II or saline | Echo, dP/dt and sacrifice |

### 9. Experimental conditions

#### 9a. Anesthesia

For subcutaneous minipump implantation and echocardiography, mice will be put under anesthesia with 3-4% isoflurane, after which anesthesia is kept at 1.5-2.5% isoflurane, as also described in SOPs and respectively. The dP/dt measurement will be performed under urethane anesthesia.

#### 9b. Pain relief

Prior to minipump implantation we will administer the pain killer carprofen (2.5mg/kg).

**9c. Euthanasia and humane end points**

After the dP/dt measurement, the mouse will be sacrificed for perfusion-fixation under urethane-anesthesia. In case of premature sacrifice of a mouse based on the criteria below (humane end points), the mouse will be euthanized with an i.p. overdose of 200mg/kg pentobarbital.

*Humane end points*

A mouse will be prematurely euthanized in case of:

- swelling and inflammation of the wound after minipump implantation, open wound with exiting of the pump and abscesses.
- Overt heart failure as a consequence of Ang II infusion, with difficulty of breathing and apathy.
- Other illnesses or conditions that induce severe pain, discomfort or suffering.

The Animal Welfare will be informed in case of calamities.

**Care****10a. Discomfort**

All animals will receive at least one tail vein injection (and max. 3) prior to the start of the experiment. At the beginning of the study (day 0) they will undergo echocardiography and minipump implantation (one anesthetization). At day of sacrifice (day 28) or the day before (day 27), all mice will first be echoed under isoflurane anesthesia. Urethane anesthesia will be administered to awake animals on day 28 for the hemodynamic measurements and sacrifice. Waking-up before urethane administration is essential for the animals to recover from the isoflurane anesthesia. Without prior recovery the drop-out numbers during the hemodynamic measurement would increase dramatically.

The overall duration of the study is 31 days for groups 1-4 and 49 days for groups 5-8. The discomfort is set to 04 (moderate-severe) because all animals will be put under anesthesia thrice.

**Group 1-4**

| type                                | severity | duration      | frequency        |
|-------------------------------------|----------|---------------|------------------|
| i.v. injection in restrainer        | 02       | < 1 minute    | daily for 3 days |
| echo (isoflurane)                   | 02       | ~15 min       | twice in 4 weeks |
| minipump implantation (isoflurane)  | 03       | ~15 min       | once             |
| hemodynamic measurements (urethane) | 02       | ~15 min       | once             |
| possible heart failure              | (04)     | unforeseeable |                  |
| possible other illnesses            | (02)     | unforeseeable |                  |

**The discomfort is estimated to sum up to max. 04 (moderate-severe)**

**Group 5-8**

| type                                | severity | duration      | frequency        |
|-------------------------------------|----------|---------------|------------------|
| i.v. injection in restrainer        | 02       | < 1 minute    | daily for 3 days |
| echo (isoflurane)                   | 02       | ~15 min       | twice in 4 weeks |
| minipump implantation (isoflurane)  | 03       | ~15 min       | once             |
| hemodynamic measurements (urethane) | 02       | ~15 min       | once             |
| possible heart failure              | (04)     | unforeseeable |                  |
| possible other illnesses            | (02)     | unforeseeable |                  |

**The discomfort is estimated to sum up to max. 04 (moderate-severe)**

**10b. Evaluation of well-being**

Angiotensin II infusion, scrambled antagomiR and AAV9 have no visible influence on the welfare of mice with this genetic background. However, we do not yet know the influence of ... manipulation, which could promote heart failure. After echo and/or minipump implantation procedures on day 0, mice will be placed in a heated room (30°C) to recover, so that they don't need to spend energy on body temperature maintenance.

### 11. Care and housing

**Groups 1-4:** The mice will be housed within the CPV. On the day of sacrifice they will be transported to the laboratories on the 4th floor.

**Groups 5-8:** The mice will be housed within the CPV in a DM-III room and on the day of experiments mice will be transported to the laboratories on the 4<sup>th</sup> floor (as approved in GGO

For all animals, water and food will be provided *ad libitum*. During the experiment the researcher will take care of the animals in consultation with veterinarians from the CPV department. In case of emergencies, the VO should be contacted.

### 12. Expertise

All interventions and procedures are performed by authorized and qualified persons (art. 9 or art. 12).

### 13 Standard Operation Procedures (SOP)

Echocardiography mouse:

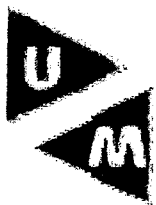
Implantation of Alzet osmotic minipumps in the mouse

Terminal hemodynamic characterization of the mouse:

### Relevant literature

Sayed D, Hong C, Chen IY, Lypowy J, Abdellatif M. *MicroRNAs play an essential role in the development of cardiac hypertrophy*. Circ Res. 100 (2007). 416-424.

Schroen B, Heymans S. *MicroRNAs and beyond: the heart reveals its treasures*. Hypertension 54 (2009). 1189-1894



University Maastricht

Faculty of Health, Medicine

and Life Sciences

Dierexperimenten Commissie

DEC

Aan:

, voorzitter  
p/a Secretariaat DEC-UM  
Postbus 616  
NL-6200 MD Maastricht  
Telefoon: 043 386 1111

Uw referentie:

Onze referentie : P

Maastricht, 28-09-2011

Geachte Onderzoeker,

Uw projectaanvraag: "*Unraveling the role of microRNA in a model of heart failure*", is op de DEC vergadering van 23 september 2011 besproken.

De DEC heeft een aantal vragen en opmerkingen:

- De duur van de proef op het voorblad is niet juist (dit is de langste periode binnen één project dat één dier in proef is). De DEC verzoekt dit aan te passen. (49 dg)
- De DEC verzoekt het GGO nummer van het voorblad te verwijderen. GGO is niet van toepassing op dit protocol (er wordt niet gewerkt met genetisch gemodificeerde dieren). *MM laten weten.*
- Bij punt 10a verzoekt de DEC de tekst om te zetten in een tabel waarin de aard, ernst, duur en frequentie van de individuele handelingen **per** groep weergegeven wordt, te sommeren tot een totaal ongerief **per** groep en tevens de laatste 2 punten van punt 9c (humane eindpunten) en het leven met "heartfailure" ook te vermelden.
- De DEC verbaast zich erover dat er geen relevante literatuur bestaat over dit onderwerp.

Gelieve eventuele vragen te beantwoorden in een brief en indien noodzakelijk Uw project aan te passen en duidelijk de aanpassingen grijs te markeren.

Uw project staat bij de DEC geregistreerd onder nummer 2011-120, gelieve dit nummer in verdere correspondentie te vermelden.

Hoogachtend,

Voorzitter DEC-UM

Maastricht, 3 oktober 2011

Betreft: DEC 2011-120

Geachte DEC,

Middels deze brief wil ik graag Uw vragen beantwoorden met betrekking tot project 2011-120, "*Unraveling the role of microRNA in a model of heart failure*". De aanpassingen zijn grijs gemarkeerd op het voorblad en in de aanvraag zelf.

- De duur van de proef op het voorblad is aangepast (49 dagen).
- Het GGO nummer is van het voorblad verwijderd.
- Bij punt 10a is de tekst omgezet in een tabel waarin de aard, ernst, duur en frequentie van de individuele handelingen per groep weergegeven wordt, alsook het opgetelde totaal ongerief per groep. Het leven met "heart failure" en mogelijke andere optredende ziekten (punt 9c, humane eindpunten) zijn ook in de tabel vermeld.
- Relevante literatuur is toegevoegd.

Ik hoop hiermee Uw vragen naar voldoening te hebben beantwoord.

Met vriendelijke groet,

Aan

Ons kenmerk

1

Doorkiesnummer

Maastricht

04-10-2011

Project: *Unraveling the role of microRNA in a model of heart failure.*

*in a model of heart*

DEC-UM

Voorzitter DEC-UM

Verantwoordelijk onderzoeker (VO):

p/a secretariaat DEC-UM

Secretariaat DEC-UM

Namens de Vergunninghouder van de DEC-UM, delen wij u mede dat voornoemd project aan de ethische toetsingscriteria voor proefdiergebruik voldoet.

Bezoekadres

De DEC maakt geen bezwaar tegen uitvoering van dit project zoals aangevraagd en geeft een **positief advies**.

Postadres

Postbus 616

6200 MD Maastricht

Projectnummer: 2011-120

Diersoort: muis

Aantal dieren: 148

Einddatum: 04-10-2015

Uw project staat bij de DEC en CPV geregistreerd onder bovenstaand nummer. Gelieve dieren, die voor dit project bestemd zijn, ook onder dit nummer aan te vragen.

Voorzitter DEC-UM

Vicevoorzitter DEC-UM